Case Report

Immunoserologically Diagnosed *Strongyloides*-Pneumonitis with Pleural Effusion and Eosinophilia

KEIKO HIDAKA¹⁾, MASATO AGETA¹⁾, HARUHIKO MARUYAMA²⁾, Takahiko KOBAYASHI²⁾, Naoto ISHIKAWA²⁾, Masaki TOMITA²⁾, Hisamitsu IDE²⁾, Hiroshi ITOH²⁾ and Yukifumi NAWA²⁾

(Accepted for publication; March 10, 1995)

Key words: Strongyloides stercoralis, pneumonitis, pleural effusion, eosinophilia, chemotherapy, albendazole

Strongyloidiasis caused by Strongyloides stercoralis is one of the commonest intestinal helminthiases in warm climates. Because of the migration route of the parasite, the respiratory tract is, next to the digestive tube, most directly and severely affected by S. stercoralis infection (Grove, 1989). While pulmonary involvement is minimal in light infections in immunocompetent hosts, severe pulmonary damage can occur either in heavy infections in competent hosts or severe, disseminated infections in compromised hosts. Therefore, respiratory symptoms are usually observed transiently in the early stage of massive infection or in the late stage of severe cases due to auto-infection and/or dissemination (Grove, 1989). In this paper we describe an unusual case of pulmonary strongyloidiasis diagnosed only by immunoserological methods. The patient suffered from pneumonitis with pleural effusion for over a month with increasing eosinophilia but never showed abdominal symptoms. Repeated faecal and sputum examinations were all negative for S. stercoralis larvae or other parasite eggs throughout the course of the disease.

Case Report

The patient was a 51 year-old house wife born and grown up in Nichinan-City, Miyazaki, Japan. She has never been abroad. On 18 August 1994, she visited a primary physician because of high fever and cough. In spite of treatment with antibiotics her symptoms turned worse and pneumonitis was suspected by chest radiogram, so that she was transferred to a regional hospital on 13 September for further work-up. On the time of admission, total white blood cell was 12310/mm³ with 21.0% eosinophils. C-reactive protein (CRP) was 14.9 mg/ dl. Other laboratory data related liver and kidney functions were within normal range. The patient was negative for antibodies to human T-cell leukemia virus type-1 (HTLV-1). Chest radiogram (Fig. 1) and computed tomogram (CT: Figs. 2-5) showed bilateral pleural effusion and multiple infiltrations predominantly in the left lung. From these observations, she was strongly suspected to be infected with helminth parasites. Because neither parasite eggs nor larvae were detected by repeated stool examinations, her serum was sent to the Department of Parasitology, Miyazaki Medical College for immunoserological examination. By a multiple-dot ELISA test, the patient's serum and pleural effusion showed positive reaction against Strongyloides ratti antigen with weak cross reaction against other nematode antigens (Fig. 7). The reaction patterns of the patient's serum and pleural effusion were essentially identical with those of the sera obtained from

¹⁾Department of Internal Medicine, Miyazaki Prefectural Nichinan Hospital, Nichinan, Miyazaki 887, Japan. ²⁾Department of Parasitology, Miyazaki Medical College,

Kiyotake, Miyazaki 889-16, Japan. 日高恵子 上田正人 (宮崎県立日南病院内科) 丸山治彦 小林隆彦 石川直人 富田雅樹 井手 久満 伊藤浩史 名和行文 (宮崎医科大学寄生虫 学教室)

177



- Fig. 1 Chest radiogram at the time of admission showing bilateral multiple diffuse infiltrations predominantly on the left lung field.
- Figs. 2–5 Computed tomogram at the time of admission showing multiple diffuse infiltrations (Figs. 2–4) and bilateral pleural effusion (Fig. 5).
- Fig. 6 Chest radiogram one month after chemotherapy. Note completely normalized appearances.



Fig. 7 Multiple-dot ELISA of the patient's serum (a) and pleural effusion (b) showing positive reaction against S. ratti antigen. Di: Dirofilaria immitis, Pw: Paragonimus westermani, Tc: Toxocara canis, Pm: Paragonimus miyazakii, Al: Ascaris lumbricoides, Fh: Fasciola hepatica, As: Anisakis simplex, Se: Spirometra erinacei, Ad: Ancylostoma duodenale, Gd: Gnathostoma doloresi, Sr: Strongyloides ratti, Ts: Trichinella spiralis

Each spot was blotted with 1 $\mu g/\mu l$ antigen (crude extract in phosphate buffered saline) and the sheet was blocked by incubating with 1% casein/Tris buffer. The sheet was incubated with 1:200 diluted patient's serum or pleural effusion at 37°C for 30 min, washed, and then incubated with peroxidase-labeled rabbit antihuman IgG at 37°C for 30 min. Peroxidase was detected using H₂O₂ and 4-chrolo-1-naphthol.

confirmed cases of strongyloidiasis (kindly supplied by Prof. Y. Sato, Department of Parasitology, the University of Ryukyus, Okinawa). She was treated successfully with albendazole (400 mg/day for 3 days from 7 October and 21 October), which was kindly supplied as an orphan drug distributed from the Project Team for the Development of the Treatment of Tropical Diseases, Ministry of Health and Welfare, Japan, through the courtesy of Prof. Y. Sato. About one month after the treatment, her symptoms completely disappeared with normalization of chest radiogram (Fig. 6) and peripheral blood cell counts (Fig. 8). During admission and subsequent follow-up study, larvae were never detected by repeated stool examinations in direct smear, AMS-III method, filter paper culture, and the most sensitive agar culture method. Nor were the larvae found in the sputum throughout the period examined. Further follow-up is now undergoing in our laboratories.

Discussion

Because of the complicated life cycle of the





parasite, strongyloidiasis is a systemic disease, though the mature adult is an intestinal nematode. Since filariform larvae migrate through the lungs as part of their life cycle, pulmonary complications should occur during the course of *S. stercoralis* infection. Strongyloidiasis is considered as an opportunistic infectious disease where the severity of the infection reflects host's immunological status. In immunocompetent hosts infestation can remain asymptomatic for years, and pulmonary complications are rather considered as the signs of overwhelming infection in the compromised host (Berger et al., 1980; Bruno et al., 1982; Shiroma et al., 1990). Surprisingly, the patient reported here had no past history of serious illness nor had been treated with immunosuppressive drugs. Furthermore, parasite larvae were never detected by repeated stool examinations during admission or subsequent follow-up studies. Therefore, it seems quite probable that the patient was immunocompetent and that she had a massive infection immediately before the respiratory symptoms had first appeared.

In Japan, the Satsunan-Islands including Amami and Ryukyu Archipelagos have been known as the heavily endemic area. Even nowadays, the prevalence of strongyloidiasis in Okinawa Prefecture, covering the Ryukyu Archipelagos, is estimated to be 5-10% among the middle and upper-age brackets of the inhabitants (Sato 1986; Shiroma et al., 1990). Although Miyazaki Prefecture was an endemic area of strongyloidiasis in the 1940s to 1950s, the prevalence at that time was less than 1% (reviewed by Tanaka, 1962) and further surveillance has never been carried out since that time. Since the patient has never been abroad nor had lived in heavily endemic areas, and since she showed no sign of immunosuppression, she was presumably infected recently somewhere in Miyazaki Prefecture. If this case were in fact caused by S. stercoralis, strongyloidiasis is still endemic at least in the southern part of Miyazaki Prefecture.

In Okinawa Prefecture over 50% of the strongyloidiasis patients were found to be infected concurrently with HTLV-1 (Nakada *et al.*, 1984) and this parasite was assumed as a leukemogenic co-factor of adult T-cell leukemia (Yamaguchi *et al.*, 1987). Although Miyazaki Prefecture, especially the area where the present patient is living, is also known as an endemic area of HTLV-1 (Tachibana *et al.*, 1984), the patient was negative for antibodies to HTLV-1, indicating that severe infection of the patient was not due to underlying concurrent infections with HTLV-1 viruses.

In the present study, the patient was diagnosed as

strongyloidiasis by immunoserological examinations and diagnostic chemotherapy. In spite of vigorous pathological changes in the lungs, the parasite larvae were never detected in the stool or sputum specimens throughout the period examined. Tanaka (1958) reported that, after percutaneous infection with the infective larvae of S. stercoralis in human. rhabditiform larvae became detectable in the stool within a month or so. The exact reason why the larvae were not detected in the stool or sputum of the patient remains unclear. One possibility is that the causative parasite was Strongyloides spp. other than S. stercoralis. If this is the case, S. fülleborni should be considered because this species was frequently found in wild Japanese monkeys in the southern part of Miyazaki Prefecture (Horii et al., 1982). Since S. ratti larval antigen was used for the dot ELISA test, further determination of the exact species of the causative parasite is necessary. Another possibility is that the patient might have an unnoticed infection with S. stercoralis in the past and became resistant to challenge infection. In experimental infection with S. ratti in rats (Moqbel, 1980), severe inflammatory response predominated by eosinophil infiltration was observed in lungs after challenge infection.

The patient was effectively cured by albendazole. Recently Shikiya *et al.* (1992) reported that, against *S. stercoralis*, ivermectin was superior to benzimidazole derivatives. At the present, both albendazole and ivermectin are available only as orphan drugs. We urge the earliest evaluation of these drugs in the chemotherapy of strongyloidiasis.

In conclusion, we demonstrated that severe pulmonary complications could occur in the early stage of strongyloidiasis in immunocompetent hosts. Physicians as well as parasitologists should be aware of such cases.

Acknowledgements

The authors wish to thank Prof. Y. Sato, Department of Parasitology, School of Medicine, The University of Ryukyus, for supplying us positive control sera and albendazole with valuable suggestions. Excellent technical assistance of Ms Ayumi Tanaka for immunoserological diagnosis is gratefully acknowledged.

References

- Berger, R., Kraman, S. and Paciotti, M. (1980): Pulmonary strongyloidiasis complicating therapy with corticosteroids. Report of a case with secondary bacterial infections. Am. J. Trop. Med. Hyg., 29, 31–34.
- Bruno, P., McAllister, K. and Matthews, J. I. (1982): Pulmonary strongyloidiasis. South. Med. J., 75, 363– 365.
- Grove, D. I. (1989): Clinical manifestations. In; Strongyloidiasis. A major roundworm infection of man. D. I. Grove ed. Taylor & Francis, London.
- Horii, Y., Imada, I., Yanagida, T., Usui, M. and Mori, A. (1982): Parasite changes and their influence on the body weight of Japanese monkeys (*Macaca fuscata*) of the Koshima troop. Primates, 23, 416–431.
- 5) Moqbel, R. (1980): Histopathological changes following primary, secondary and repeated infections of rats with *Strongyloides ratti*, with special reference to tissue eosinophils. Parasite Immunol., 2, 11–27.
- Nakada, K., Kohakura, M., Komoda, H. and Hinuma, Y. (1984): High incidence of HTLV antibody in carriers of *Strongyloides stercoralis*. Lancet i, 633.
- Sato, Y. (1986): Epidemiology of strongyloidiasis in Okinawa. In: Collected Papers on the Control of Soiltransmitted Helminthiasis, Vol. 3, Yokogawa, M. et al.,

ed., The Asian Parasite Control Organization, Tokyo, 20–31.

- Shikiya, K., Kinjo, N., Uehara, T., Uechi, H., Ohshiro, J., Arakaki, T., Kinjo, F., Saito, A., Iju, M. and Kobari, K. (1992): Efficacy of ivermectin against *Strongyloides stercoralis* in humans. Int. Med., 31, 310–312.
- Shiroma, Y., Kiyuna, S. and Sato, Y. (1990): Clinical studies on human strongyloidiasis in Okinawa, Japan. Jpn. J. Parasitol., 39, 277–283.
- 10) Tachibana, N., Okayama, A., Kusune, E., Yokota, T., Shishime, E., Tsuda, K. and Matano, H. (1984): Antihuman T-cell leukemia virus antibody distribution in Miyazaki Prefecture. J. Jpn. Assoc. Infect. Dis., 58, 717–722 (in Japanese with English abstract).
- Tanaka, H. (1958): Experimental and epidemiological studies on strongyloidiasis of Amami Oshima Island. Jpn. J. Exp. Med., 28, 159–182.
- 12) Tanaka, H. (1962): On the genus *Strongyloides*. In; Prog. Med. Parasitol. Jpn. (Japanese edn.), Vol. 2, pp. 241–277. K. Morishita, Y. Komiya and H. Matsubayashi eds. Meguro Parasite Museum, Tokyo (in Japanese).
- 13) Yamaguchi, K., Matutes, E., Catovsky, D., Galton, D.A.G., Nakada, K. and Takatsuki, K. (1987): *Strongyloides stercoralis* as candidate co-factor for HTLV-I induced leukaemogenesis. Lancet ii, 94–95.