Research Note

Development of *Babesia gibsoni* in the Midgut of the Tick, Haemaphysalis longicornis

SEIICHI HIGUCHI, SUMITO SIMOMURA, SEIICHI KAWAMURA AND YOSHIO YASUDA

(Accepted for publication; October 2, 1990)

Key words: Babesia gibsoni, Haemaphysalis longicornis, development

Although the development of *Babesia* spp. in the tick midgut has been described by a number of investigators (Higuchi *et al.*, 1989; Mehlhorn *et al.*, 1980; Mehlhorn and Walldorf, 1988; Shortt, 1973; Stewart, 1978; Stewart *et al.*, 1986), detailed observations on the development of *B.gibsoni* (Patton, 1910) in its ixodid host are lacking.

This paper describes the morphological changes observed during the development of B.gibsoni in Haemaphysalis longicornis, the tick vector. Haemaphysalis ticks, removed from a local cow, were bred and maintained in the laboratory on two male New Zealand White rabbits which were 8 and 11 months old and weighed 2.42 and 2.65 kg, respectively. The adult ticks were infected with *B.gibsoni* by permitting them to feed on splenectomized, infected dogs. The strain of B.gibsoni used had been isolated from naturally infected dogs in Towada area Aomori Prefecture, Japan (Itoh et al., 1987). Five splenectomized and infected mongrel dogs, 6 to 12 months old, were used for infecting ticks. A total of 240 infected ticks were examined, during the nymphal stage and after moulting to adults in an incubator maintained at 25°C and 80% relative humidity. The ticks were teased apart in a Ringer's solution formulated for insects, under a dissecting microscope. Each midgut was stained with Giemsa's stain prior to microscope examination.

By 6 hr post-engorgement (PE), merozoites of B.gibsoni were observed free of erythrocytes in the midgut contents of the ticks (Fig. 1). Within 24 hr PE, relatively large round-forms $2-3 \mu m$ in diameter, the so-called "ring-forms", were detected (Fig. 2). In the ring-forms, the nucleus was located in the ring and the cytoplasm was basophilic. Soon after, the rings developed into spherical-forms which were $3-4 \,\mu m$ in diameter. These spherical-forms had an eosinophilic nucleus and a light basophilic cytoplasm (Fig. 3). Within 2–4 days PE, large bizarre-forms (6–7 μ m in diameter) were found (Fig. 4). During this stage some of the bizarre-forms elongated into cylindrical forms $6-8 \ \mu m$ in length (Fig. 5). Within 5-6 days PE, even larger round or ellipitical forms (7–9 μ m in diameter) were observed in the gut (Fig. 6). The nucleus was peripheral and cytoplasm stained light blue with Giemsa. About 7 days PE, the large globular organisms gradually began to decrease in number, and finally disappeared from the gut. Within 24 hr PE, concurrent with the appearance of the rings forms, was the disappearance of most of the ingested erythrocytes. This may have been due to red cell hemolysis caused by enzymes from disrupted eosinophil granules during the process of phagocytosis (Schleger, 1976).

The spherical forms, which appeared in the midgut during the first 24 hr of engorgement,

Department of Veterinary Internal Medicine School of Veterinary Medicine and Animal Sciences, Kitasato University, Towada, Aomori 034, Japan 樋口誠一 下村澄人 川村清市 安田純夫(北里 大学獣医畜産学部)



Fig. 1. The merozoite of *B. gibsoni* in the tick midgut $\times 2,000$.



Fig. 4. The so-called "bizarre"-form, in the gut $\times 2,000$.



Fig. 2. The round "ring-form" in gut contents $\times 2,000$.



Fig. 3. Rings developing into spherical-forms within the gut $\times 2,000$.



Fig. 5. Elongated organisms in the gut $\times 2,000$.



Fig. 6. A Round-form, possibly a zygote, in the gut $\times 2,000$.

494

likewise have been reported in ticks infected with B.canis (Mehlhorn et al., 1980), B.ovata (Higuchi et al., 1989), B.bovis (Stewart, 1978), b.caballi (Holbrook et al., 1968), and B.bigemina (Stewart et al., 1986). The B.gibsoni spherical forms were smaller than *B.canis* and *B.bigemina*, but similar in size to B.ovata, B.caballi, and B.bovis (Table 1). The so-called "bizarre" forms of B.gibsoni, which we observed in the midgut within 24 hr PE, have been shown to occur during the life cycle of the other Babesia species as well (Higuchi et al., 1989; Mehlhorn et al., 1980; Shortt, 1973; Stewart, 1978; Stewart et al., 1986). Within 2-4 days PE, elongated forms of the parasite then appeared. The microgametes of B.gibsoni were difficult to identify. It was considered, however, that the spherical and elongated forms might be macrogametes and microgametes, respectively, based on their morphological characteristics, the time of their appearance in the gut lumen of the tick, and by comparison with the other Babesia species. Within 5-6 days PE, large globularshaped organisms, 7.0–9.0 μ m in diameter, appeared. These forms have also been reported in the tick phase of the life cycle of B.ovata and B.argentina (Riek, 1966), now synonymous with B.bovis (Hoyte, 1976). Though the process of fusion of the macrogametes was not observed in this study, the round forms of the parasite have been identified as zygotes by other workers (Riek, 1966; Schein et al., 1975). About 7 days engorgement, round, putative zygotes gradually dis-

TIME POSTINFE	CTION*	BABESIA SP AND (TICK HOST)			
	B.gibsoni	B. canis	B. ovata	B. bigemina	B. bovis
	(Haemaphysalis) longicornis	(Dermacentor) reticulatus	(Haemaphysalis) longicornis	(Boophilus) microplus)	(Boophilus) microplus)
Within 12 hours	ring-forms (2~3 μm)	spherical- stages (6~7 μm)	ring-forms (2~3 μm)	large spherical- forms	binary- fission
12~24 hours	spherical- forms (3~4 μm)	polymorphous stages (5 \sim 6 μ m)	ring and spherical- forms (4~5μm)	fission- body (20 μm)	spherical- forms
2~4 days	bizarre- forms (6~7μm)	spindle- shaped stages $(6 \sim 8 \mu m)$	fission-forms (4 ~5 μm)	immature- fission body	elongated- forms
	elongated- forms (6~8 μ m)	(bizarre- forms	spherical- forms	large vermicules
4~5 days	zygotes (7~9μm)	polymorphous stages (6∼7µm)	elongated- forms (6∼8µm)	elongated organisms	
5~6 days		slender stages (10~12μm	round-forms) (9~10 μm)		
6~8 days					
8~12 days			vermicules forms (13~15 μm)		

Table 1 Comparison of developmental stages of Babesia species in ticks*

* Data from: B. canis: Mehlhorn et al. (1980); B. ovata: Higuchi et al. (1989); B. bigemina: Stewart et al. (1986); B. bovis: Stewart (1978). appeared from the midgut of the tick.

Table 1 compares the developmental characteristics of *B.gibsoni* with other *Babesia* species, in the tick. The morphology of *B.gibsoni* shows a close similarity to *B.canis, B.bigemina* and *B.ovata*. The timing of vermicle maturation varies with the *Babesia* species (Higuchi et al., 1989; Mehlhorm et al., 1980; Stewart, 1978; Stewart et al., 1986). Differences in timing of stages of the several *Babesia* species may be due to differences between the various strains and



Fig. 7. Schematic diagram of the development of B. gibsoni in the midgut of the tick, H. longicornis.
a: merozoites, b: ring-form, c: spherical-form, d: bizarre-form, e: elongated-form, f: round-form. Refer to the foot-notes of Fig. 1 to 6.

f

species of the tick hoste. Based on the present findings, the development of *B.gibsoni in the midgut of the tick H.longicornis*, is diagrammed in Fig. 7.

References

- Higuchi, S., Ezura, K., Hamana, M., Kawamura, S. and Yasuda, Y. (1989): Development of *Babesia* ovata in the midgut of the tick, *Haemaphysalis* longicornis. Jpn. J. Vet. Sci., 51, 1129–1135.
- Holbrook, A. A., Anthony, D. W. and Johnson, A. J. (1968): Observations on the development of Babesia caballi (Nuttall) in the tropical horse tick Dermacentor nitens Neumann. J. Protozool., 15, 391-396.
- Hoyte, H. M. D. (1976): The tick-fever parasites of cattle. Proc. R. Soc. Qd., 87, 5–13.
- Itoh, N., Higuchi, S., Ogasawara, T., Ogasawara, A. and Kawamura, S. (1980): An outbreak of canine babesiosis in Aomori prefecture. J. Jpn. Vet. Med. Assoc., 40, 167–171 (In Japanese).
- Mehlhorn, H., Schein, E. and Voigt, W. P. (1980): Light and electron microscopic study on developmental stages of *Babesia canis* within the gut of the tick *Dermacentor reticulatus*. J. Parasitol., 66, 220-228.
- Mehlhorn, H. and Walldorf, V. (1988): Life cycles. In Parasitology in Focus, Ed. Mehlhorn, H., Springer-Verlag Berlin Hideberg New York London Paris Tokyo, 1–48.
- Patton, W. S. (1910): Preliminary report on a new *Piroplasma (Piroplasma gibsoni* sp. nov.) found in the blood of the madras hunt and subsequently discovered in the blood of the jackal *Canis aureus*. Bull. Soc. Pathol. exot., 3, 274–281.
- Riek, R. F. (1966): The life cycle of Babesia argentina (Lignieres, 1903) (Sporozoa: Piroplasmidea) in the tick vector Boophilus microplus (Canestrini). Aust. J. Agric. Res., 17, 247-245.
- Schein, E., Buscher, G. and Friedhoff, K. T. (1975): Lichtmikroskopische Untersuchungen uber die Untwicklung von *Theileria annulata* in *Hyalomma anatolicum exacavatum*. I. Die Entwicklung im Darm vollgesogener Nymphen. Z. Parasitenkd., 48, 123–136.
- Schleger, A. V. (1976): Boophilus microplus: Cellular responses to larval attachment and their relationship to host resistances. Aust. J. Bio. Sci., 29, 499-512.
- Shortt, H. E. (1973): *Babesia canis*: The life cycle and laboratory maintenance in its arthropod and mamalian host. Int. J. Parasitol., 3, 119–148.
- 12) Stewart, N. P. (1978): Differences in the life cycles between a vaccine strain and an unmodified strain

496

of *Babesia bovis* (Babes, 1989) in the tick *Boophilus* microplus (Canetrini). J. Protozool., 25, 497–501.

 Stewart, N. P., Dalgliesh, R. J. and de Vos, A. J. (1986): Effect of different methods of maintenance on the development and morphology of *Babesia* bigemina in the gut of *Boophilus microplus*. Res. Vet. Sci., 40, 94–98.